

The effect of *Lactobacillus acidophilus* on the clearance of pneumococcus pathogen from mice lung

Haider Yousif AL-Murayati

Department of Biology, College of Science, University of Al- Mustansiriyah

Abstract:

In the present study, we investigate the preventive effect of intraperitoneal inoculation of *Lactobacillus acidophilus* against intranasally challenge with *Streptococcus pneumonia* using a mouse experimental model.

Five days before challenge with Capsulated *S. pneumonia* the treated group injected intraperitoneally with 0.5 ml of heat –killed *Lactobacillus acidophilus*, while the positive control group injected with 0.5 ml of phosphate buffer saline. Mice were infected by dropping 25 µl of the inoculums containing 10^8 log- phase colony forming unit *S.pneumonia* in PBS into each nostril and allowing it to be inhaled by holding the mice in head-up vertical position for 2 min . Animals were sacrificed by cervical dislocation on day 1st, 2^{ed} and 3rd day of infections.

The study showed that numbers of pneumococcal pathogen that recovered from lungs of positive control mice was increased from the first day of infection (Log 5.7 ± 1.19) and the increasing rate continued in the next 2 days (Log 6.9 ± 1.06 at 3rd day). In contrast the animal treated with *L. acidophilus* showed greater resistance to pneumococcal challenge, as their pathogen count in lung was significantly lower than those in positive control at same intervals (on day one was Log 2.5 ± 0.65 to end as Log 2 ± 0.43 at third day), likewise the log mean number of bacteria found in blood stream of untreated animal increased steadily over the 3 days of the experiments starting with Log 3.3 ± 0.94 to Log 4.96 ± 0.7 at the last day, while the treated mice showed significant low level of Bacteremia gradually reaching to Log 0.5 ± 0.1 at the last, also the challenge with *S. pneumonia* increased the number of blood leukocytes in both experimental group of our study, but the pretreated mice with *L. acidophilus* present a significant higher value than positive control group. Also, the neutrophils were the principle cell population responsible for the increasing leukocyte of both study groups, followed by lymphocytes cells.

These results showed that administration of intraperitoneal *L. acidophilus* may increases the clearance rate of *S. pneumoniae* from lung and prevent the dissemination of pneumococci into blood and that may due to activation of immune system by increasing the total count and efficacy of leukocytes especially neutrophils and lymphocytes.

الخلاصة:

تم التحري عن التأثير الوقائي للتلقيح داخل الغشاء البريتوني لبكتريا *Lactobacillus acidophilus* ضد اصابات الجهاز التنفسي الناتجة عن بكتريا *Streptococcus pneumonia*. حققت فئران المجموعة الاختبارية لمدة خمسة ايام بـ 0.5 مل من عالق بكتريا الـ *L. acidophilus* المقتولة بالحرارة، وحققت مجموعة السيطرة الموجبة بـ 0.5 مل من دارئ الفوسفات. أحدثت الأصابة ببكتريا *S.pneumonia* بتقطير 25µl من العالق البكتيري (الحاوي على 10^8 من الخلايا المكونة للمستعمرات في الطور اللوغارتمي) في كل فتحة من فتحات الأنف ولتسهيل استنشاق اللقاح تم مسك الفئران بشكل عمودي لمدة دقيقتان، قتلت الفئران في الأيام 1, 2, 3 بواسطة الخلع العنقي (cervical dislocation). أظهرت نتائج هذه الدراسة أن أعداد بكتريا الـ *S. pneumonia* المسترجعة من انسجة الرئة لحيونات السيطرة الموجبة زادت منذ اليوم الاول (Log 5.7 ± 1.19) للاصابة، واستمرت الزيادة بالتقدم خلال اليومين اللاحقين حتى وصلت الى (Log 6.9 ± 1.06) في اليوم الثالث، وعلى العكس من ذلك فإن الحيوانات التي عوملت ببكتريا الـ *L. acidophilus* أظهرت مقاومة ملحوظة للاصابة بالبكتريا المرضية وان تعداد البكتريا في الرئة كان منخفضا بصورة معنوية عن حيوانات السيطرة الموجبة (Log 2 ± 0.43 في اليوم الاول الى Log 2.7 ± 0.65 في اليوم الثالث).

كذلك كان لو غارتم معدل الاعداد للبكتريا المسترجعة من الدم لحيوانات السيطرة متزايدا بصورة متواترة خلال الثلاثة ايام من التجربة ابتداء بـ $\text{Log } 3.3 \pm 0.94$ وأنتهاء بـ $\text{Log } 4.96 \pm 0.7$ في اليوم الثالث، فيما اظهرت الحيوانات المعاملة مستويات معنوية واطنة من جرثوم الدم وبشكل تدريجي لتبلغ $\text{Log } 0.5 \pm 0.1$ في اليوم الاخير. لوحظ من خلال النتائج أن الإصابة بـ *S. pneumoniae* أدت الى زيادة في عدد كريات الدم البيضاء لكلا المجموعتين. لكن الحيوانات المعاملة بـ *L. acidophilus* أظهرت زيادة معنوية عالية مقارنة بحيوانات السيطرة الموجبة. وجد من خلال العد التفريقي أن كريات الدم العدلة (Neutrophils) هي المسؤل الاول عن هذه الزيادة في كلا المجموعتين وتلتها في العدد كريات الدم اللمفاوية (Lymphocytes). أشارت النتائج الى أن التلقيح داخل الغشاء البريتوني ببكتريا الـ *L. acidophilus* ممكن أن يزيد من معدل إزالة بكتريا *S. pneumoniae* من الرئة ويمكن ايضا أن يمنعها من الأنتشار الى الدم، وقد يعزى ذلك الى تعزيز دور الجهاز المناعي بزيادة اعداد كريات الدم العدلة واللمفاوية وزيادة فعاليتها.

Introduction:

The upper airways of humans (nose and pharynx) may harbor potentially pathogenic bacteria (PPB) (*Staphylococcus aureus* is one of the most important, but *Streptococcus pneumoniae*, β -hemolytic streptococci, and *Haemophilus influenzae* are also members of this group).

In patients carrying PPB, antiseptic regimens could be crucial for infection control after major operations or injuries of the head, nasal sinuses, or lungs. Such regimens may also be important for diabetic patients and persons receiving hemodialysis, in intensive care units, or with impaired immunity due to various other causes^[1].

The widespread antibiotics usage exerts a selective pressure that acts as a driving force in the development of antibiotics resistance^[2].

A multi-resistant strain becomes a serious global problem, which constrains scientists to search for new effective therapeutic agents. Alternative therapeutic options should be used as strategies to prevent the selective development of antibiotic resistant bacterial strains, restore a balanced microbial flora and enhance the defense mechanisms of the human body. These criteria are best fulfilled by live microorganisms which are naturally hosted by the human body^[3].

The idea that guts microflora play a very important role in the maintenance of health and well-being is acquiring worldwide acceptance^[4]. Lactic acid bacteria (LAB) are present in the intestine of most animals and the beneficial role

played by these microorganisms in humans and other animals has been reported extensively^[5,6].

Recently LAB have become an attractive option of modern medical practice and the attention has been paid to their health-promoting properties of particular importance are their probiotics properties and specially the ability to compete with pathogens in vivo^[7].

The host defense against PPB is mainly the task of the immune system. In mammals, the mucosal surface area represents an extensive interface with the external environment through which pathogens mainly initiate infections. The mucosal surfaces of the upper airways are well equipped with an immune system that reacts to some extent independently of the immune system, and these surfaces are functionally linked to other mucosal surfaces^[8].

Lactobacilli can directly promote the development of the gut mucosal barrier by the stimulation of the immune response. Since immune mechanisms at different mucosal sites interact through the common mucosal immune system, LAB could also play an important role in disseminating effector cells to lung and other mucosal sites^[9]. Clinical trials have demonstrated that probiotics may decrease the incidence of respiratory tract infections^[10].

In this study we evaluate the role of intraperitoneal injection of heat killed *L. acidophilus* as a probiotic bacteria on clearance rate of *S. pneumoniae* from mice lung and dissemination of pneumococcal in the blood.

Material and Methods:

Animals:

Male 6 week-old Swiss blab\c mice were obtained from Pharmaceutical Control department of the Ministry of Health, they were housed in plastic cages at room temperature and fed mice chow and water ad libitum. Mice divided into 3 groups consisting of 5 mice, in addition to the 3 positive control groups^[11].

- 1 - *Lactobacillus acidophilus* (bacteria were obtained from department of biology college of science, AL-Mustansiriya university), and cultured on De man Rogsa sharp medium (MRS) at 37°C for 48hrs.washed with distilled water ,killed at 100C° for 30 min, and suspended in phosphate buffer saline (PBS) at desired concentration just before use.
- 2 - Capsulated *S. pneumonia* was isolated from respiratory tract of patient in central child hood hospital, Baghdad. *S. pneumonia* was first grown on blood agar for 18 hrs; freshly grown colonies were suspended in brain hart infusion broth and incubated at 37°C overnight. The pathogens harvested through centrifugation at 3000 rpm for 10 min and then washed 3 times with sterile PBS. The infecting dose was chosen according to Julio *et al.*^[9].

Experimental infection:

For five days before challenge with *S. pneumonia*, 0.5 ml of heat-killed LBA was injected intraperitoneally (IP)^[11].

Mice were infected by dropping 25µl of the inoculum containing 10⁸ Log-phase colony forming unit(CFU)of *S.pneumonia* in PBS into each nostril and allowing it to be inhaled by holding the mice in head-up vertical position for 2 min^[9]. Mice were killed on day 1st, 2^{ed}, and 3rd day of infection.

Determination of bacterial growth in lung homogenates and blood:

S. pneumonia was challenged to positive control mice (non-treated) and mice that had been treated with *L. acidophilus*. After challenge, mice were

killed by cervical dislocation, lung were excised to determine the bacterial count by homogenization in 1ml sterile normal saline and plated in duplicate on blood agar. Bacteraemia was monitored by sampling the blood via cardiac puncture by a heparinized syringe and cultured in duplicate on blood agar. Result was reported as Log CFU/ml after 18h and 37C° incubation period as negative or positive haemocultured^[12].

Determination of total and differential numbers of leukocytes in the blood:

The total numbers of leukocytes was determined with haemocytometer and differential cell count was performed by counting 200 cells in blood smears with Giemsa stain^[13].

Statistical examination:

Results were expressed as the mean ± standard deviation (SD). The paired t-test was used to test for differences between the control group and the treated group. Significant difference was defined as $P < 0.05$.

Results and Discussion:

Probiotics are live microorganisms thought to be healthy for the host organism. According to the currently adopted definition by FAO/WHO, probiotics are: "Live microorganisms which when administered in adequate amounts confer a health benefit on the host"^[14].

The use of probiotics in the treatment of specific diseases has evolved into an extremely valuable option yet to be optimally used in clinical medicine^[15].

This study concern on the protective effect of heat-killed *L. acidophilus* against *S.pneumonia* infections in male swiss mice with age of 6 weeks was studied. The heat-killed *L.acidophilus* was injected intraperitoneally for five days before challenge with vital *S. pneumonia*.

The results showed that numbers of pneumococcal pathogen that recovered from lungs of positive control mice at the 1st day of infection was Log 5.7±1.19, the

increasing rate continued in the next 2 days to reach $\text{Log } 6.9 \pm 1.06$ at 3rd day. In contrast the animal treated with *L. acidophilus* showed greater resistance to pneumococcal challenge, as their pathogen

count in lung was significantly lower than those in positive control at same intervals (on day one was $\text{Log } 2.5 \pm 0.65$ to end as $\text{Log } 2 \pm 0.43$ at third day (Fig-1).

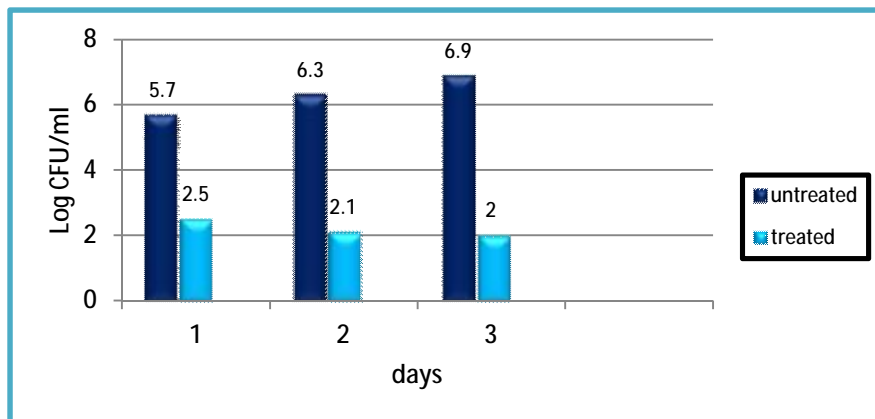


Fig-1: Log mean numbers of bacteria that recovered from mice lungs

The Log mean number of bacteria found in blood stream of untreated animal was increased steadily over the 3 days of the experiments starting with $\text{Log } 3.3 \pm$

0.94 to $\text{Log } 4.96 \pm 0.7$ at the last day, while the treated mice showed significant low level of Bacteremia gradually reaching to $\text{Log } 0.5 \pm 0.1$ at the last day (Fig-2).

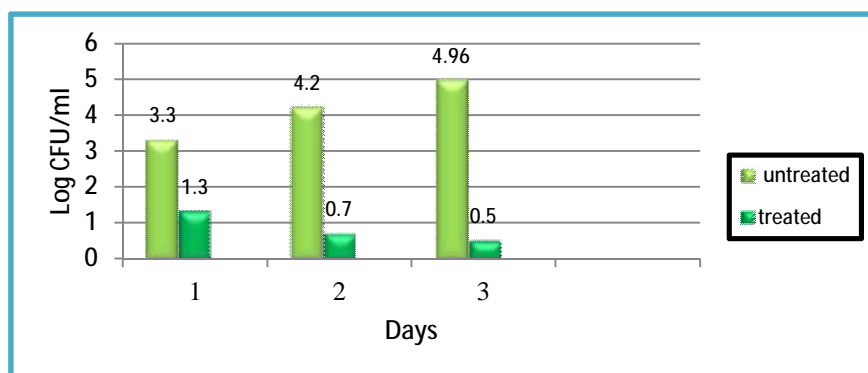


Fig-2: Log mean numbers of bacteria recovered from mice blood.

These finding are in agreement with Julio *et al*^[16], they found that pneumococcal colonization of lung and bacteremia were significantly greater in control group comparison with *L. casei* orally pretreated group, they found that the number of bacteria in lung and blood tended to decrease ($P < 0.05$) during infection period and they suggest that the addition of *L. casei* to the diet has a beneficial effect because it accelerate the recovery of the innate immune response

and improve the specific mechanisms against *S. pneumonia* respiratory infection in malnourished mice.

Other studies^[9] found that the oral administration of yoghurt contains two type of lactic acid bacteria (*L. bulgaricus*, *L. thermophilus*) induced an early recovery of immunological parameters and accelerate the normalization of immune response against the pathogenic respiratory agents while Alvarez *et al*^[17] stated that the oral administration of *L.asei* or yoghurt

to young mice enhanced lung clearance of *pseudomonas aeruginosa*.

Nasally administration of live or heat killed *L. casei* prevent the dissemination of the pathogenic bacteria to the blood and induced its lung clearance [18]. Rosa *et al.* [19] demonstrated that intranasal administration of *L. fermentum* isolated from pharynx of BALB/C mice was able to reduce pathogen count from all respiratory tract organs and that indicates the presence of Lactobacilli lead to decrease the adherence or faster clearance of *S. pneumonia* from respiratory tract. Marcela *et al.* [20] evaluate the effect of nasal administration of *L. lactis* to Swiss albino mice and showed that such treatment was able to increase the clearance rate of *S. pneumonia* from lung

and prevent spreading the pathogen to the blood

and they suggest that the effect induced by the nasal inoculation of *L.lactis* could be explained by a decreased adherence of *S.pneumoniae* to the respiratory epithelium, as it has been reported that nasally given LAB could exclude competitively pneumococcal cells [21] Saito *et al.* [22] refer to a single intraperitoneal injection of *L.casei* into mice led to nonspecific resistance against intraperitoneal challenge with lethal dose of *pseudomonas aeruginosa*.

The challenge with *S. pneumonia* increased the number of blood leukocytes in both experimental group of our study, but the pretreated mice with *L. acidophilus* present a significant higher value (P<0.05) than positive control group (Fig-3).

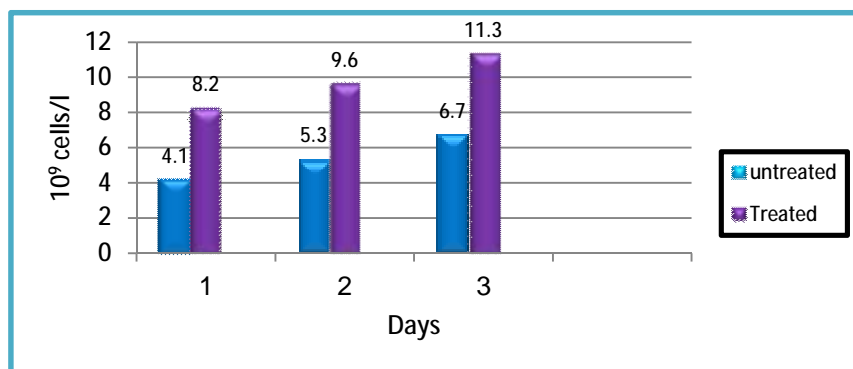
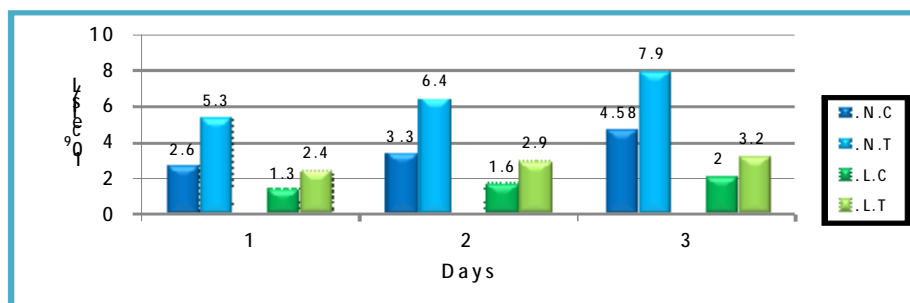


Fig-3: Total WBC count in positive control group (untreated) and treated mice

The neutrophil was the principle population responsible for the increasing

leukocyte of both study groups, followed by lymphocytes cells (Fig-4).



- N.C.=Neutrophils of positive control mice
- N.T. = Neutrophils of pretreated mice
- L.C. = Lymphocytes of positive control mice
- L.T. = Lymphocytes of pretreated mice

Fig-4: Neutrophils and lymphocytes counts of positive control and pretreated mice

In an uninfected lung, neutrophils are in the blood (with increased numbers marginated within the pulmonary capillaries compared with other blood vessels) but not the air spaces^[23]. However, within hours of infection, neutrophils begin to appear in the interstitial compartments and air spaces of the lungs^[24, 25].

Several studies demonstrated that phagocytic and microbial activity of neutrophils in blood and those recruited into the lung are great importance for the control of pneumococcal infections^[26,27]. Also alveolar macrophage constitute the first line of phagocytic defense against infectious agent that evade the mechanical defense and gain access to the gas-exchange air ways^[28], these macrophage are capable of generating numerous mediators that induce the recruitment of neutrophils from pulmonary vasculature into the alveolar space. These recruited neutrophils provide auxiliary phagocytic capacities that are critical for the effective eradication of offending pathogens^[28, 29, 30].

Our findings are goes with Julio *et al.*^[18], they found the nasal administration of *L. casei* to malnourished mice increase the number of leukocytes and the peroxides activity of phagocytes cells in the blood and that would explain why low or no bacteria were detected in blood, they suggest that the treatment with probiotic bacteria exerts effects on hematopoiesis. Another study^[16] showed that a progressive increase in white blood cells was observed in mice that receive orally *L. casie* in their diet after infection with *S. pneumonia* and the differential count indicate that this increasing phenomenon involved mainly neutrophil and they found that infection rapidly triggered the neutrophil recruitment into the alveoli of mice resulting in increased cell counts in broncho alveolar lavage (BAL). Also Salva *et al.*^[31] reported that mice replete with *L. casei* and challenged with *S. pneumonia* had an elevated level of blood neutrophil and this elevation continue until the 5th day post infection.

Marcela *et al.*^[20] refered to an increase in activation of macrophage observed in mice treated with *L. lactis* by nasal administration and that enhance the protective effect, this activation would induce more effective neutrophil recruitment^[32,33]. Hidemura *et al.*^[34] stated that impaired neutrophil recruitment into local inflammatory site can be restored by administration of *Bifidobacterium longum* as a probiotic.

The protective effect of treatment with *Lactobacillus* against the pneumococcal colonization of lung, bacteremia and lung tissue injury was correlated with the stimulation of the systemic and respiratory immune response^[18].

Pulmonary lymphocytes play an important role in lung defense against infection because they influence important events of inflammation and tissue repair, such as the recruitment of phagocytes from blood and fibroblast activation^[35]. Salva *et al.*^[31] demonstrated that the use of *L. casei* CRL 431, as a supplement in a repletion diet, induced an improvement in the production and maturation of myeloid and lymphoid cells. Perdigon *et al.*^[36] showed that feeding with milk fermented with *L. casei*, *L. acidophilus* or a mixture of both produces a remarkable effect on the immunostimulation in the host and he found that macrophage and lymphocyte activation was much higher in cultures fermented with *L. casei* and *L. acidophilus* or both than in the controls or in the individual or mixed culture without a fermentation process. Such enhancement of the immune response might be due to the substances produced by these organisms during the fermentation process, such as some metabolites and casein peptides, bacterial enzymes (proteolytic and acteriolytic) that improve the digestibility of the milk constituents.

Conclusion:

These results from this study suggest that the previous intraperitoneal administration of heat-killed *L.*

acidophilus to pneumococcal infected mice has a beneficial effect because it reduce significantly the colonization of pneumococcus pathogen in lung also increase the number of leukocytes mainly neutrophil and that may enhance the resistance of respiratory tract against the *S. pneumonia* infection.

Further experiments are required to establish the mechanism by which *L. acidophilus* isolate affects on *L. acidophilus* pathogenicity. In the future, the immunological aspects of the protective role of *L. acidophilus* should be studied.

References:

- 1- Ulrich Glück and Jan-Olaf Gebbers. Ingested probiotics reduce nasal colonization with pathogenic bacteria (*Staphylococcus aureus*, *Streptococcus pneumoniae*, and β -hemolytic streptococci). American Journal of Clinical Nutrition, 2003. Vol. 77 (2). Pp: 517-520.
- 2- Beovic, B. The issue of antimicrobial resistance in human medicine. Intern. J. of Food Microbiology. 2006. Vol. 112. Pp: 280–287.
- 3- Rayes, N.; Seehofer, D.; Muller A. R.; Hansen, S.; Bengmark, S. and Neuhaus, P. Influence of probiotics and fibre on the incidence of bacterial infections following major abdominal surgery. Z. Gastroenterol. 2002. Vol. 40. Pp: 869–76.
- 4- Hentges, D. J. Gut flora and disease resistance. Fuller, R. eds. Probiotics: The Scientific Basis Chapman and Hall London, UK. 1992. Pp: 87-110
- 5- Cross, M. L. Immunoregulation by probiotic lactobacilli: pro Th1 signals and their relevance to human health. Clin. Appl. Immunol. Rev. 2002. Vol. 3. Pp: 115-125.
- 6- Perdigón, G.; Fuller, R. and Raya, R. Lactic acid bacteria and their effect on the immune system. Curr. Issues Intest. Microbiol. 2001. Vol. 2. Pp: 27-42.
- 7- Petrova, M.; Petrova, M.; Georgieva, R.; Dojchinovska, L.; Kirilov; N.; Iliev, I.; Antonova, S.; Hadjieva, N.; Ivanova, I and Danova, S. Lactic acid bacteria against pathogenic microbes. Trakia. J. of Sciences, 2009. Vol. 7 (2) Pp: 33-39.
- 8- Glück, U. Antisepsis of the nasal cavity. In: Kramer A, Heeg P, Botzendhard K, eds. Hygiene in hospital and practice. Munich, Germany: Urban & Fischer Verlag. 2001. Pp: 260–4.
- 9- Julio Villena; Silvia Racedo; Graciela Agüero and Susana Alvarez. Yoghurt accelerates the recovery of defence mechanisms against *Streptococcus pneumonia* in protein-malnourished mice. British Journal of Nutrition. 2006. Vol. 95 (3) Pp: 591-602.
- 10- Katja Hatakka; Erkki avilahti; Antti Pönkä; Jukka, H.; Meurman; Tuija oussa; Leena Näse; Maija Saxelin; and Riitta Korpela. Effect of long term consumption of probiotic milk on infections in children attending day care centers: double blind, randomized trial. BMJ. 2. 2001. Vol. 322 (7298). Pp: 1327.
- 11- Miake, S.; Nomoto, K.; Yokokura. T.; Yoshikai, Y.; Mutai, M. and Nomoto, K. Protective effect of *Lactobacillus casei* on *Pseudomonas aeruginosa* infection in mice. Infect. Immun. 1985. Vol. 48 Pp: 480-485.
- 12- Andrea Marra and Daniel Brigham. *Streptococcus pneumoniae* causes experimental meningitis following intranasal and otitis media infections via a nonhematogenous route .infect Immun. 2001. Vol. 69 (12) Pp: 7318–7325.
- 13- Bergeron, Y.; Oullet, N.; Deslauriers, A.; Simard, M.; Olivier, M. and Bergeron, M. G. Cytokine kinetics and other host factors in response to pneumococcal pulmonary infection in

- mice. *Infect. Immun.* 1998. Vol. 66. Pp: 912-922.
- 14- Report of a Joint FAO- WHO expert consultation on evaluation of health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria. 2001 .
- 15- Vesterlund, S.; Karp, M.; Salminen, S. and Arthur, C. *Staphylococcus aureus* adheres to human intestinal mucus but can be displaced by certain lactic acid bacteria. *Microbiology.* 2006. Vol. 152 Pp: 1819-1826.
- 16- Julio Villena; Silvia Racedo; Graciela Agüero; Elena Bru; Marcela Medina and Susana Alvarez. *Lactobacillus casei* Improves Resistance to Pneumococcal Respiratory Infection in Malnourished Mice. *J. Nutr.* 2005. Vol. 135. Pp: 1462-1469.
- 17- Alvarez, S.; Herrero, C.; Bru, E. and Perdigo'n, G. Effect of *Lactobacillus casei* and yogurt administration on prevention of *Pseudomonas aeruginosa* infection in young mice. *J Food Prot.* 2001. Vol. 64. Pp: 1768–1774.
- 18- Julio Villena; Natalia Barbieri; Susana Salva; Mat'ias Herrera and Susana Alvarez Enhanced immune response to pneumococcal infection in malnourished mice nasally treated with heat-killed *Lactobacillus casei*. *Microbiol Immunol.* 2009. Vol. 53. Pp: 636–646.
- 19- Rosa Cangemi de Gutierrez a; Viviana Santos a and Maria Elena Nader-Macias Protective effect of intranasally inoculated *Lactobacillus fermentum* against *Streptococcus pneumoniae* challenge on the mouse respiratory tract. *Immunology and Medical Microbiology.* 2001. Vol. 31. Pp: 187-195.
- 20- Marcela Medina; Julio Villena; Susana Salva; Elisa Vintin; Philippe Langella and Susana Alvarez. Nasal administration of *Lactococcus lactis* improves local and systemic immune responses against *Streptococcus pneumoniae*. *Microbiol Immunol.* 2008. Vol. 52. Pp: 399–409.
- 21- Fowler, N. G. Antimicrobials and competitive exclusion. *Int. J. Food Microbiol.* 1992. Vol. 15 Pp: 277–9.
- 22- Saito, H.; Watanabe, T. and Horikawa, Y. Effects of *Lactobacillus casei* on *Pseudomonas aeruginosa* infection in normal and dexamethasone-treated mice. *Microbiol. Immunol.* 1986. Vol. 30 (3). Pp: 249-59.
- 23- Doerschuk, C. M. Mechanisms of leukocyte sequestration in inflamed lungs. *Microcirculation.* 2001. Vol. 8: Pp: 71–88.
- 24- Burns, A. R.; Smith, C.W. and Walker, D.C. Unique structural features that influence neutrophil emigration into the lung. *Physiol Rev.* 2003. Vol. 83 Pp: 309–336.
- 25- Mizgerd, J. P. Molecular mechanisms of neutrophil recruit-ment elicited by bacteria in the lungs. *Semin Immunol* 2002. Vol. 14. Pp: 123–132.
- 26- Bergeron, Y.; Ouellet, N.; Deslauriers, A. M.; Simard, M.; Olivier, M. and Bergeron M.G. Cytokine kinetics and other host factors in response to pneumococcal pulmonary infection in mice. *Infect Immun.* 1998. Vol. 66. Pp: 912–22.
- 27- Wang, E.; Simard, M.; Ouellet, N.; Bergeron Y.; Beauchamp D. and Bergeron M.G. Pathogenesis of pneumococcal pneumonia in cyclophosphamide induced leukopenia in mice. *Infect Immun.* 2002. Vol. 70. Pp: 4226–38.
- 28- Sibille, Y. and Reynolds, H.Y. Macrophages and polymorpho-nuclear neutrophils in lung defense and injury. *Am. Rev. Respir. Dis.* 1990. Vol. 141. Pp: 471–501.
- 29- Gingles, N. A.; Alexander, J. E. Kadioglu, A.; Andrew, P. W.; Kerr, A.; Mitchell, T. J.; Hopes, E.; Denny, P.; Brown, S.; Jones, H. B.; Little, S.; Booth, G.C. and McPheat, W.L. Role of genetic resistance in invasive

- pneumococcal infection: identification and study of susceptibility and resistance in inbred mouse strains. *Infect Immun* 2001. Vol.69. Pp: 426–434.
- 30- Gordon, S. B.; Irving, G. R.; Lawson, R. A.; Lee M.E. and Read, R.C. Intracellular trafficking and killing of *Streptococcus pneumoniae* by human alveolar macrophages are influenced by opsonins. *Infect Immun*. 2000. Vol. 68 Pp: 2286–2293.
- 31- Salva, S.; Villena, J.; Racedo, S.; Agüero, G. and Alvarez, S. *Lactobacillus casei* addition to a repletion diet induced early normalization of cytokine profiles during pneumococcal infection in malnourished mice. *Food Agric Immunol*. 2008. Vol. 19. Pp: 195–211.
- 32- Standiford, T. J.; Kunkel, S. L.; Greenberger, M. J.; Laichalk, L. L. and Strieter, R. M. Expression and regulation of chemokines in bacterial pneumonia. *J. Leukoc. Biol*. 1996. Vol. 59. Pp: 24–8.
- 33- Nelson, S. and Summer, W.R. Innate immunity, cytokines and pulmonary host defense. *Infect Dis Clin North Am*. 1998. Vol. 12. Pp: 555–67.
- 34- Hidemura, A.; Saito, H.; Fakatsu, K.; Matsuda, T.; Kitayama, J.; Ikeda, S.; Kang, W. and Nagawa, H. Oral administration of *Bifido-bacterium longum* culture condensate in a diet-restricted murine peritonitis model enhances polymorphonuclear neutrophil recruitment into the local inflammatory site. *Nutrition*. 2003. Vol .19. Pp: 270–274.
- 35- Furfaro, S.; Berman, J. S. and Center, D. M. Determinants of T cell migration in the lung. In: Kradin R.L., Robinson B.W.S., eds. *Immuno-pathology of Lung Disease*. Boston, N. J.: Butterworth Heinemann, 1996. Pp: 15–32.
- 36- Perdigon, G.; de Macías, M. E.; Alvarez, S.; Oliver, G. and Holgado, A. P. Systemic augmentation of the immune response in mice by feeding fermented milks with *Lactobacillus casei* and *Lactobacillus acidophilus*. *Immunology*. 1998. Vol. 63. Pp: 17–23.