#### Serum Leptin levels in Obese Post Menopause Women

#### Al-Zubaidy A. Ghaith\*, Fadhil A. Khazraj \* Department of Pharmacotherapeutics, College of Pharmacy, University of Al-Mustansiriyah

الخلاصة

الهدف من هذه الدراسة هو إيجاد العلاقة بين هرمون النحافة وحالة السمنة في النساء ما بعد سن اليأس.

تم قياس مستوى هرمون النحافة في مصل الدم بطريقة الامتزاز المناعي المرتبط بانزيم. كذلك تم قياس منسب كتلة الجسم باستخدام الوزن (كغم)/المتر المربع, وقياس محيط الخصر بالسنتيمترات.

تم اخذ هذه المعايير من 100 سيدة ما بعد سن اليأس والتي قسمت تبعا للسمنة الى مجموعتين: 60 سيدة مصابة بالسمنة و 40 سيدة غير مصابين بالسمنة.

لقد وجد فرق معنوي عالي في مستويات هرمون النحافة في مصل الدم ومنسب كتلة الجسم ومحيط الخصر في النساء المصابات بالسمنة ما بعد سن اليأس مقارنة بالنساء الغير مصابات بالسمنة (p<0.01).

كما وجد ارتباط عالي بين هرمون النحافة و منسب كتلة الجسم (r=0.739, p≤0.01) وكذلك بين هرمون النحافة ومحيط الخصر (r=0.684, p≤0.01).

#### Abstract

The objective of this study is to find the relationship between leptin and obesity in post menopause women. Serum leptin level was determined by using the Enzyme Linked Immuno Sorbant Assay (ELISA) technique, The Body Mass Index (BMI) was measured by weight (kg)/square height (m<sup>2</sup>), and waist circumference was measured in centimeters.

These parameters where taken from 100 post menopause women who were divided according to BMI into two groups: 60 obese women and 40 non-obese women.

A highly significant difference in Serum Leptin levels, BMI and Waist circumference was found in the obese post menopause women compared with non-obese subjects (p $\leq$ 0.01). And a highly significant positive correlation between serum leptin levels and BMI (r=0.739, p $\leq$ 0.01), and between serum leptin levels and waist circumference (r=0.684, p $\leq$ 0.01).

## Introduction

Leptin was discovered as a result of studies on ob/ob mice, a strain of hyperphagic obese mice that were known to lose weight when their circulation was attached to normal mice (parabiosis)<sup>[1]</sup>. Subsequent studies revealed that ob/ob mice had a mutation that results in inability to produce a protein, first called the ob protein and later Leptin, which regulates food intake <sup>[2]</sup>. In addition to being very obese, these mice grew poorly and had infertility due to gonadal hypofunction. Administration of Leptin to these animals resulted in a marked decrease in food intake, weight loss, and improved growth <sup>[3]</sup>.

Leptin is a member of the cytokine family, and its receptor is a member of the gp130 group of cytokine receptors, there are at least five forms of the Leptin receptor <sup>[4]</sup>. The most widely distributed is the short form of the receptor, which is present in most tissues and may serve to transport Leptin into the brain. The long form of the receptor is located in areas where Leptin is thought to act, including hypothalamic nuclei. There may also be a circulating form of the Leptin <sup>[4]</sup>.

Circulating factors that bind Leptin might also contribute to Leptin resistance <sup>[5]</sup>. In one study, C-reactive protein was identified as a circulating factor that binds to Leptin, impairs its signaling, and attenuates its physiologic effects (in cultured cells and an ob/ob mouse model). In addition, physiologic concentrations of Leptin stimulated C-reactive protein expression in vitro <sup>[5]</sup>.

Food intake is reduced by systemic Leptin administration in normalweight experimental animals, but the response decreases as the animals become obese. However, when Leptin is injected into the ventricular system of the brain of obese animals, they remain responsive <sup>[6]</sup>. Since Leptin is transported across the blood-brain barrier to act within the brain, the processes controlling the entrance of Leptin into the brain are pivotal determinants of its action on food intake <sup>[7]</sup>.

Most obese people, however, are not Leptin deficient, and serum Leptin concentrations are directly related to their amount of body fat. In several surveys of obese subjects, no mutations in the Leptin gene were detected <sup>[8, 9]</sup>. Given the high serum Leptin concentrations and apparent Leptin resistance in obese subjects <sup>[10, 11]</sup>, little response to exogenous Leptin might be expected. However, in a study of the effect of recombinant Leptin (0.01, 0.03, 0.1, or 0.3 mg/kg per day) or placebo in normal-weight subjects for four weeks and obese subjects for 24 weeks, there was a very modest dose-dependent decrease in weight in both groups <sup>[12]</sup>. After weight loss, Leptin administration prevents the decline in metabolic rate and circulating concentrations of thyroid hormone <sup>[13]</sup>.

Postmenopausal women are at increased risk of coronary heart disease (CHD), partly because of the decline in estrogen production and concurrent elevations in total and low-density lipoprotein (LDL) cholesterol level <sup>[14]</sup>. Obesity, weight gain, and adverse changes in body fat distribution and composition are part of this phenomenon <sup>[15, 16]</sup>. Moreover, the rise in LDL

cholesterol levels and onset of other CHD risk factors (e.g. high blood pressure, high total cholesterol and triglyceride levels, and insulin resistance) is directly influenced by weight gain <sup>[16,17]</sup>.

Several longitudinal studies also suggest that menopause increases central adiposity, although these studies used waist circumference or waist-hip ratio (WHR), a less precise technique <sup>[15, 18]</sup>.

This present study was designed to evaluate the role of leptin level in obese post menopause women.

## **Materials and Methods**

## Subjects:

This study included 100 post menopause women from the external laboratory department at Baghdad teaching hospital. Subjects recruited to full fill the criteria of being postmenopausal (at least 6-months history of amenorrhea not due to pregnancy and age range 47-66 years).

Study subjects were divided according to obesity into two groups:

Group 1: 60postmenopausal obese women (body mass index  $\geq$ 30 kg/m<sup>2</sup>).

Group 2: 40 postmenopausal non-obese women (body mass index <30 kg/m<sup>2</sup>). **Methods:** 

# Serum leptin:

For each women included in this study venous blood samples were collected to obtain the serum. The Leptin (sandwich) Enzyme immunoassay kit provides materials for the quantitative determination of leptin in serum and plasma. This assay is intended for in vitro diagnostic use only. The leptin (sandwich) ELISA is a solid phase enzyme-linked immunosorbent assay (ELISA) based on the sandwich principle.

The microtiter wells are coated with a monoclonal antibody directed towards a unique antigenic site on a Leptin molecule. An aliquot of patient sample containing endogenous Leptin is incubated in the coated well with a specific rabbit anti Leptin antibody. A sandwich complex is formed. After incubation the unbound material is washed off and an anti rabbit peroxidase conjugate is added for detection of the bound Leptin. Having added the substrate solution, the intensity of color developed is proportional to the concentration of Leptin in the patient sample, reads at 450nm with the microtiterplate reader within 10 minutes after adding the stop solution Normal value in Female = 7.36  $\pm 3.73$  ng/ml and in Male =  $3.84 \pm 1.79$  ng/ml.<sup>[19]</sup>.

#### Anthropometric measures:

1- Body Mass Index (BMI) was calculated by weight (in kilograms) divided by the square of height (in meters), weight and height are measured by the same scale for the all sample subjects.

BMI = Weight (kg)/Square Height (m<sup>2</sup>)<sup>[20]</sup>.

2 - Waist circumference was measured in centimeters (cm) using a flexible nonelastic measuring tape.

#### **Statistics:**

To compare the significance of the difference in the mean values of any two groups, Student's t-test was applied;  $p \le 0.01$  was considered statistically highly significant,  $p \le 0.05$  was considered statistically significant.

The Pearson correlation coefficient [r] test is used to describe the association between the different studied parameters;  $p \le 0.01$  was considered statistically highly significant,  $p \le 0.05$  was considered statistically significant.

#### **Results:**

There was no significant difference in mean age between obese and nonobese subjects (p = 0.341) (table 1). Body mass index was higher in obese subjects than non-obese subjects (p  $\leq$  0.01) (table 1). Mean waist circumference was higher in obese group than non-obese group (p  $\leq$  0.01) (table 1). Mean serum Leptin level was higher in obese group than non-obese group (p  $\leq$ 0.01) (table 1).

Characteristic	Obese group	Non-obese group	p value
Age (year)	$55.87 \pm 4.38$	$56.65 \pm 3.34$	0.341
BMI (kg/m <sup>2</sup> )	$35.78 \pm 5.06$	$25.94 \pm 3.22$	≤0.01
WC (cm)	$112.5 \pm 8.6$	85.54 ± 17.16	≤0.01
Leptin (ng/ml)	$31.41 \pm 4.79$	$17.3 \pm 5.7$	≤0.01

# Table 1: Mean $\pm$ SD values of age, body mass index, waist circumference & Leptin in obese (n = 60) and non-obese (n = 40) subjects.

Mean waist circumference correlates positively with body mass index [r = 0.759, p  $\leq$  0.01] (figure 1). Serum Leptin correlates positively and strongly with body mass index [r = 0.739, p  $\leq$  0.01] (figure 2). Serum Leptin also correlates positively with waist circumference [r = 0.68, p  $\leq$  0.01] (figure 3).



Figure 1: Correlation between waist circumference and body mass index.



Figure 2: Correlation between serum Leptin and body mass index.



Figure 3: Correlation between serum Leptin and waist circumference.

## Discussion

Serum leptin concentration is increased in obese subjects and is closely related to fat mass and BMI <sup>[21, 22]</sup>. It is regulated by serum insulin concentration<sup>[23]</sup> and declines with weight loss <sup>[21]</sup>. Several reports have shown a higher leptin concentration in women than in men <sup>[24]</sup>. The gender difference has been explained partly by the variable degree and distribution of the amount of body fat depots. Women tend to have a higher overall obesity which is more pronounced in subcutaneous fat than in visceral fat, in contrast to men who have a lower overall but greater visceral adiposity. Serum leptin concentrations are not influenced by menopausal status or serum estradiol level <sup>[25]</sup>.

In the present study of the relationship between serum Leptin and obesity in postmenopausal women, we found that mean serum Leptin level is higher in obese group than non-obese group (Mean  $\pm$  SD;  $31.41 \pm 4.79$  vs.  $17.3 \pm 5.7$ ng/ml; p $\leq$ 0.01), also Serum Leptin correlates positively and strongly with body mass index [r = 0.739, p  $\leq$  0.01], similar correlation is present in a study done by Marita A.R. et. al. (2005) <sup>[26]</sup>. In addition, a previous study reported in a crosssectional study of 3,553 subjects in Netherlands, that BMI and waist circumference are positively associated with serum leptin concentration <sup>[27]</sup>, this is also true in regard for the relation between serum leptin level and waist circumference [r = 0.68, p  $\leq$  0.01], another study reported that women who are overweight or had a higher waist circumference (women  $\geq$  88 cm) have a significantly higher risk of having hyperleptinemia <sup>[28]</sup>.

Other studies concluded that fat distribution contributes to the variability in serum leptin in obese patients. In particular, subcutaneous abdominal fat is a determinant of leptin concentration, also independently of the amount of fat mass, whereas the contribution of preperitoneal visceral fat is not significant <sup>[29]</sup>.

Most obese individuals are leptin-resistant <sup>[30]</sup>, resistance to the actions of leptin could be cause by decreased leptin transport through the blood-brain barrier <sup>[31, 32]</sup>, or to reduced signaling distal to the leptin receptor <sup>[31, 33]</sup>. Peripheral signals such as gloucocorticoids may also interfere with leptin's interaction with its receptor and produce central leptin resistance <sup>[34, 35]</sup>.

The mechanism underlying the elevated circulating levels of leptin on obese women, may be due to an accelerated secretion rates of the peptide from adipose tissue because of increased ob gene expression <sup>[36]</sup>. In addition, subcutaneous fat produces more leptin mRNA than visceral fat, which could explain why women have higher leptin levels in as much as they have more subcutaneous fat than visceral fat <sup>[37]</sup>.

# References

- Coleman DL. Effects of parabiosis of obese with diabetes and normal mice. Diabetologia 1973; 9:294.
- 2 Zhang Y, Proenca R, Maffei M, et al. Positional cloning of the mouse obese gene and its human homologue. Nature 1994; 372:425-32.
- 3 Pelleymounter MA, Cullen MJ, Baker MB, et al. Effects of the obese gene product on body weight regulation in ob/ob mice. Science 1995; 269:540-3.
- 4 Tartaglia LA, Dembski M, Weng X, et al. Identification and expression cloning of a leptin receptor, OB-R. Cell 1995; 83:1263-71.
- 5 Chen K, Li F, Li J, et al. Induction of leptin resistance through direct interaction of C-reactive protein with leptin. Nat Med 2006; 12:425-32.
- 6 Van Heek M, Compton DS, France CF, et al. Diet-induced obese mice develop peripheral, but not central, resistance to leptin. J Clin Invest 1997; 99:385-90.
- 7 Banks WA, Kastin AJ, Huang W, et al. Leptin enters the brain by a saturable system independent of insulin. Peptides 1996; 17:305-11.
- 8 Considine RV, Considine EL, Williams CJ, et al. Evidence against either a premature stop codon or the absence of obese gene mRNA in human obesity. J Clin Invest 1995; 95:2986-8.
- 9 Maffei M, Stoffel M, Barone M, et al. Absence of mutations in the human OB gene in obese/diabetic subjects. Diabetes 1996; 45:679-82.
- 10 Saad MF, Riad-Gabriel MG, Khan A, et al. Diurnal and ultradian rhythmicity of plasma leptin: effects of gender and adiposity. J Clin Endocrinol Metab 1998; 83:453-9.
- 11 Kennedy A, Gettys TW, Watson P, et al. The metabolic significance of leptin in humans: gender-based differences in relationship to adiposity, insulin sensitivity, and energy expenditure. J Clin Endocrinol Metab 1997; 82:1293-1300.

- 12 Heymsfield SB, Greenberg AS, Fujioka K, et al. Recombinant leptin for weight loss in obese and lean adults: a randomized, controlled, dose-escalation trial. JAMA 1999; 282:1568-75.
- 13 Rosenbaum M, Murphy EM, Heymsfield SB, et al. Low dose leptin administration reverses effects of sustained weight-reduction on energy expenditure and circulating concentrations of thyroid hormones. J Clin Endocrinol Metab 2002; 87:2391-4.
- 14 Matthews KA, Meilahn E, Kuller LH, et al. Menopause and risk factors for coronary heart disease. N Engl J Med 1989; 321(10):641-6.
- 15 Poehlman ET, Toth MJ, Gardner AW. Changes in energy balance and body composition at menopause: a controlled longitudinal study. Ann Intern Med 1995; 123(9):673-5.
- 16 Wing RR, Matthews KA, Kuller LH, et al. Weight gain at the time of menopause. Arch Intern Med 1991; 151(1):97-102.
- 17 Denke MA, Sempos CT, Grundy SM. Excess body weight. An underrecognized contributor to dyslipidemia in white American women. Arch Intern Med 1994; 154(4):401-10.
- 18 Tchernof A, Poehlman ET. Effects of the menopause transition on body fatness and body fat distribution. Obes Res 1998; 6(3):246-54.
- 19 Goldsby R. A., Kindt T. J. and Osborne B. A. Ku By Immunology. Fourth edition. Printed in USA. (2000):161-169.
- 20 Garrow JS, Webster J: Quetelet's index (W/H2) as a measure of fatness. Int J Obes 1985; 9:147-153.
- 21 Considine RV, Sinha MK, Heiman ML, et al. Serum immunoreactiveleptin concentrations in normal-weight and obese humans. N Engl J Med 1996; 334:292-5.
- 22 Rosenbaum M, Nicolson M, Hirsch J, Heyms SB, Gallagher D, Chu F et al. Effects of gender, body composition, and menopause on plasma concentrations of leptin. Journal of Clinical Endocrinology and Metabolism 1996; 81: 3424-3427.
- 23 Segal KR, Landt M, Klein S. Relationship between insulin sensitivity and plasma leptin concentration in lean and obese men. Diabetes 1996; 45: 988-991.
- 24 Niskanen LK, Haffner S, Karhunen LJ, Turpeinen AK, Miettinen H, Uusitupa MIJ. Serum leptin in obesity is related to gender and body fat topography but does not predict successful weight loss. European Journal of Endocrinology 1997; 137: 61-67.
- 25 Hadji P, Hars O, Bock K, et al. The influence of menopause and body mass index on serum leptin concentration. Europ J Endocr 2000; 143: 55-60.
- 26 Marita AR, Sarkar JA, Rane S. Type 2 diabetes in non-obese Indian subjects is associated with reduced leptin levels: Study from Mumbai, Western India. Mol Cell Biochem 2005; 275: 143-151.

- 27 Ruige JB, Mooy J, Dekker JM, et al. Leptin and variables of body adiposity, energy balance, and insulin resistance in a population-based study. Diabetes Care 1999; 22(7): 1097-1104.
- 28 Mendoza-Núñez VM, Garcia-Sánchez A, Sánchez-Rodriguez A, et al. Overweight, waist circumference, age, gender, and insulin resistance as risk factors for hyperleptinemia. Obes Res 2002;10(4): 253-259.
- 29 Minocci A, Savia G, Lucantoni R, et al. Leptin plasma concentrations are dependent on body fat distribution in obese patients. Intern J Obes 2000; 24: 1139-1144.
- 30 Emanuelli B, Peraldi P, Filloux C, et al. SOCS-3 inhibits insulin signaling and is up-regulated in response to tumor necrosis factor-alpha in the adipose tissue of obese mice. J Biol Chem 2001; 276: 47944-9.
- 31 Ur E, Grossman A, Despress JP. Obesity results as a consequence of glucocorticoid induced leptin resistance. Horm Metab Res 1996; 28: 744-7.
- 32 Schawatz MW, Peskind E, Raskind M, et al. Cerebrospinal fluid leptin levels: relation to plasma levels and to adiposity in human. Nat Med 1996;2: 589-93.
- 33 Frilhbecu G, Salvador J. Relation between leptin and the regulation glucose metabolism. Diabetologia 2000; 43: 3-12.
- 34 Zakrzewska KE, Gusin I, Sainsbury A, et al. Glucocorticoids as counterregulatory hormones of leptin: toward to understanding of leptin resistance. Diabetes 1997; 46: 717-9.
- 35 Kiess W, Englaro P, Hanitsch S, et al. High leptin concentrations in serum of very obese children are further stimulated by dexamethasone. Horm Met Res 1996; 28: 708-1.
- 36 Lönnqvist F, Nordfors L, Jansson M, et al. Leptin secretion from adipose tissue in women, relationship to plasma levels and gene expression. J Clin Invest 1997; 99(10): 2398-2404.
- 37 Lönnqvist F, Arner P, Nordfors L, et al. Overexpression of the obese (OB) gene in adipose tissue of human obese subjects. Nature Med 1995; 1:950-953.