Evaluating the Proposed Mechanism by Which Atorvastatin Can Protect Renal Tissue against Contrast Media: An Animal study


*Department of Pharmacology and Toxicology, College of Pharmacy, Mustansiriyah University, M.Sc. program, Iraq.
**Department of Anatomy and Histology, College of Medicine, Mustansiriyah University, Baghdad- Iraq.
***Department of Clinical Laboratory Sciences, College of Pharmacy, Mustansiriyah University, Baghdad- Iraq.

Abstract:
Contrast-induced nephropathy (CIN) is an elevation of serum creatinine of ≥ 0.5 mg/dL from baseline after two to three days of exposure to contrast substance if there is no other cause for acute kidney injury. Atorvastatin may protect normal kidney physiology from contrast-induced kidney injury by effects unrelated to hypolipidemia termed pleiotropic effect by decline of endothelin production, angiotensin system down regulation, and under expression of endothelial adhesion molecules. This study was conducted to assess the strategy by which atorvastatin can achieve protective effect for kidneys after exposure to contrast media in an animal model. A 40 male rats were distributed randomly into 4 groups; ten rats for each: group (1): given normal saline; group (2): CIN group given iopromide as contrast media; group (3): given atorvastatin (20mg/kg) and iopromide; and group (4): given atorvastatin (40mg/kg) and iopromide. Blood collected by cardiac puncture for detection of serum glutathione, malondialdehyde, matrix metalloproteinase-9, and interleukin-18. The results have shown a significant increase in inflammatory and oxidative stress markers in contrast media group, and significant reduction in these markers in atorvastatin treated groups, in a dose-dependent manner. As conclusion, atorvastatin mechanism for protection against CIN in a dose-dependent manner can mediate by anti-inflammatory and antioxidant effects.

Key words: contrast induced nephropathy, atorvastatin, oxidative stress, inflammation.
Introduction:
Contrast media (CM) is an agent that is used to enhance visibility of body tissues, usually used for blood vessels and GI organs [1]. Different types of contrast media are used in several techniques according to imaging devices, where they are used in [2]: X-ray technique; magnetic resonance imaging (MRI) signal enhancing; and ultrasound scattering. Contrast media may be negative or positive, this difference is related to absorbing fewer or more of the radiation than do the neighboring tissues, respectively [3].

Iodinated contrast agent is a form of intravenous radiocontrast having iodine, which improves the visibility of structures and organs during radiographic procedures. Usually, an iodinated contrast agent may either be oil-based or water-soluble. The oil-based is slowly absorbed by body tissue and, it is usually only used in sialographic and hysterosalpingographic studies. Water-soluble iodinated medium, which is more rapidly absorbed, may be used in place of barium sulfate for gastrointestinal examination that is contraindicated by the use of barium [4]. Iopromide is used as a water-soluble x-ray contrast agent. It is low osmolar, non-ionic contrast agent for intravascular use. It makes opaque when passing in the vessels under x-ray path, permitting imaging of internal tissues and structures. The levels of opaqueness caused by iodine materials is related directly to the quantity of iodine in contrast media (volume and concentration) [5].

Fatal anaphylactoid or life-threatening reactions may occur throughout or after iopromide administration. These responses range from itching, urticaria, bronchospasm, to facial and laryngeal edema [6]. Iodinated contrast agent has a toxic effect on the kidneys, particularly when given by arteries, during coronary angiography [7]. Contrast-induced nephropathy (CIN) is an elevation of serum creatinine of ≥ 0.5 mg/dL or ≥ 25% from baseline after two to three days of exposure to contrast substance if there is no other cause for acute kidney injury [8,9].

The pathophysiology of CIN not completely understood yet. Many issues may play an important role in the mechanism of this type of kidney damage.
These factors include indirect insult, e.g. increase renal vasoconstriction, depletion of prostaglandin formation and decrease nitric oxide; also involve the direct injury on renal tubules by free radicals' generation, more oxygen consumption, and increase internal pressure because of tubular obstruction. All these situations end with ischemia of medullary region of the kidney\textsuperscript{[10]}.

Many risk factors may enhance the attack with CIN; some of them related to patient and other with procedure itself such as types of contrast media, dose and multiple contrast media administration within 72 hours. Patient related risk factors are more important and involve: aging; heart failure; hypotension; hyperglycemia; hypo-albuminemia; renal impairment; nephro-toxic agents; and others\textsuperscript{[11]}. As prevention strategies against CIN, intravenous extracellular volume expansion with saline, using lowest dose of contrast media, and administering low osmolar non-ionic contrast media instead of high osmolar ionic type reported some effectiveness\textsuperscript{[12]}.

Like other statins, atorvastatin is a competitive inhibitor of 3-hydroxy-3-methyl-glutaryl-coenzyme A reductase inhibitor. It's used in the prevention of cardiovascular diseases in elderly patients, with high risk for coronary heart diseases, such as hypertension, age ≥ 55 years, low HDL- cholesterol, smoking, or history of family coronary heart disease.\textsuperscript{[13]}

Atorvastatin have "pleiotropic effects" antioxidant properties, by reducing reactive oxygen species (ROS) generation and lipid peroxidation. This effect mediated by decrease production of important isoprenoids intermediates (geranylgeranyl pyrophosphate and farnesyl pyrophosphate)\textsuperscript{[14]}. The anti-inflammatory effect of atorvastatin in rheumatoid conditions mediated by inhibitory effect on several proinflammatory cytokines such as interleukin-1β (IL-1β), interleukin-6 (IL-6), tumor necrosis factor-α (TNF-α), and chemokines (CCL2, CCL5) which were reduced significantly in treated arthritic rats by atorvastatin\textsuperscript{[15]}.

This study was conducted to assess the mechanism by which atorvastatin can achieve protective effect against kidney damage after exposure to contrast media in an animal model.

**Materials and Methods**

**Animals**

Forty male rats (200-300g) were involved in this study and placed in animal house in College of Pharmacy/Mustansiriyah University, feed on pellets and free water. With animals placed in cages each plastic cage (20x25x35cm) contains 3 rats. Animals left a week for environmental acclimatization, temperature (23±3 ºC), 12/12 hr light/dark rhythm. The study approved from ethics and scientific committee/College of Pharmacy/ Mustansiriyah University, this experiment was start.

**Experimental Protocol**

Rats distributed randomly to 4 groups, ten for each:

- Group (1): given saline orally for 6 days, as control.
- Group (2): CIN group; exposed to dehydration 3 days, in 4\textsuperscript{th} day given furosemide (10mg/kg) as intramuscular, 20 min later iopromide 370 mg I/ml given IV through tail vein as 10 ml/kg\textsuperscript{[16]}.
- Group (3): small-dose atorvastatin group; given atorvastatin (20mg/kg)\textsuperscript{[17]} over 6 consecutive days, exposed to dehydration between day 2 to 4, in 5\textsuperscript{th} day furosemide (10mg/kg) given through IM, 20 min later iopromide 370 mg I/ml given IV via tail vein as 10 ml/kg.
- Group (4): large-dose atorvastatin group; like with third group but atorvastatin given in 40 mg/kg.

Regarding atorvastatin preparation, a 40 mg of calcium salt as white crystalline powder was dissolved in 5 ml DW to obtain oral suspension (8 mg/ml).
Collection of Samples
Within the 7th day and after anaesthetizing each animal by ketamine and xylazine \[^{[18]}\], we collect blood specimen via intracardiac aspiration, each sample was transferred to plastic centrifuge tube and left at room temperature for complete clotting of blood. Serum was aspirated after centrifugation at 1000 rpm for 10 minutes and kept in the freeze to be ready for doing assay of serum glutathione, malondialdehyde, matrix metalloproteinase-9, and interleukin-18.

Assay of Biomarkers
Detection of serum glutathione (GSH), malondialdehyde (MDA), matrix metalloproteinase-9 (MMP-9), and interleukin-18 (IL-18) were measured by sandwich ELISA technique according to the manufacturer's instructions (Mybiosource-USA) \[^{[19]}\].

Statistical Analysis
Data were expressed as mean ± SD and the ANOVA test was used for statistical evaluation of significant difference among groups and compared with that of control, using SPSS-16 software program. P-value of < 0.5 was regarded as significant.

Results
Effect of Atorvastatin on Oxidative Stress Markers After Contrasting Induced Nephropathy
Regarding oxidative stress markers, GSH levels reduced significantly in CIN group compared with other groups \( (p < 0.05) \). Low dose atorvastatin group elevated GSH levels significantly compared with CIN group, but still significantly lower than control group and high dose atorvastatin group \( (p < 0.05) \). High dose atorvastatin significantly increased GSH levels compared with low dose atorvastatin and CIN groups \( (p < 0.05) \) and approach to control group \( (p = 0.664) \), as shown figure (1).

Considering serum MDA marker, its levels increased significantly in CIN group compared with other groups \( (p < 0.05) \). Low dose atorvastatin group reduced MDA levels significantly compared with CIN group, but still significantly higher than control group and high dose atorvastatin group \( (p < 0.05) \). High dose atorvastatin significantly reduced MDA levels compared with low dose atorvastatin and CIN groups \( (p < 0.05) \) and approach to control group \( (p = 0.926) \), as shown figure (2).

![Figure (1): Effect of atorvastatin on serum GSH levels after CIN.](image)

Data were expressed as M±SEM. Different lower-case letters indicate significant difference among groups.
**Figure (2): Effect of Atorvastatin on Serum MDA Levels After CIN.**

Data were expressed as M±SEM.
Different lower-case letters indicate significant difference among groups.

*P*-value < 0.05 considered significant difference.

MDA= Malondialdehyde, CIN= contrast induced nephropathy.

**Effect of Atorvastatin on Inflammatory Markers After Contrasting Induced Nephropathy**

Regarding inflammatory markers, IL-18 levels increased significantly in CIN group compared with other groups (*p* < 0.05). Low dose atorvastatin group reduced IL-18 levels significantly compared with CIN group, but still significantly higher than control group and high dose atorvastatin group (*p* < 0.05). High dose atorvastatin significantly reduced IL-18 levels compared with low dose atorvastatin and CIN groups (*p* < 0.05) and approach to control group (*p* =0.477), as shown in figure (3).

Considering MMP-9 marker, its level increased significantly in CIN group compared with other groups (*p* < 0.05). Low dose atorvastatin group reduced MMP-9 levels significantly compared with CIN group, but still significantly higher than control group and high dose atorvastatin group (*p* <0.05). High dose atorvastatin significantly reduced MMP-9 levels compared with low dose atorvastatin and CIN groups (*p* < 0.05) and approach to control group (*p* =0.868), as shown figure (4).
Figure (3): Effect of Atorvastatin on IL-18 Levels After CIN.
Data were expressed as M±SEM.
Different lower-case letters indicate significant difference among groups.
*P*-value < 0.05 considered significant difference.
IL-18= interleukin-18, CIN= contrast induced nephropathy.

Figure (4): Effect of Atorvastatin on MMP-9 Levels After CIN.
Data were expressed as M±SEM.
Different lower-case letters indicate significant difference among groups.
*P*-value < 0.05 considered significant difference.
MMP-9= Matrix Metalloproteinase-9, CIN= contrast induced nephropathy.

**Discussion**
In normal physiological conditions, tubular transport related to ROS generation, mostly in thick ascending limb, where high dense mitochondrial residents, which represents main sources for hydroxyl...
readicals (OH⁻) and superoxide anions (O₂⁻) generation by NADPH- oxidase. In medullary tubular levels, ROS formation has a central role in regulating renal microcirculation, mediated its effect on NO bioavailability [20].

Contrast induced nephropathy results from combination of toxic renal parenchymal injury and hypoxia. Parenchymal injury result from ROS generation and renal hypoxia induced after iodinated contrast media administration increasing ROS generation in kidney [21].

Contrast media administration significantly decrease medullary oxygenation, without reduction in tubules reabsorption. As the result, neurohumoral vasoconstrictive changes mediated by increasing the release of endothelin and prostaglandin from endothelial cells when exposed to contrast media. All these lead to oxygen imbalance and reduces mitochondrial scavengers' activity and enhance ROS generation. During hypoxia conditions, high amount of ATP converted to ADP, AMP and adenosine which is further hydrolyzed to inosine by 5-nucleotidase and then metabolized to the hypoxanthine that generates hydrogen peroxide (H₂O₂) and xanthine by xanthine oxidase enzyme. Hydrogen peroxide cause further damage to renal medullary microcirculation by scavenging NO. Also, superoxide redials reduced NO bioavailability by formation peroxynitrite, so in normal physiological conditions, NO protect against ROS-mediated endothelial injury and diminished transport-dependent ROS generation in the thick ascending limbs [21].

Previous studies have shown that oxidative stress and generation of free radicals after exposure to contrast media have an important role in pathogenesis of CIN and reduced antioxidant enzymes activity (i.e. shift the balance from antioxidant toward the oxidative stress) [22,23].

In the current study, serum GSH levels significantly decreased after exposure to contrast media, while treated atorvastatin groups show significant rise in GSH levels in a dose-dependent manner by 32% and 66.6% in low and high dose respectively, compared with CIN group. These results were similar to the Al-Otaibi et al findings (2012), who found that levels of GSH in renal tissues markedly decreased after exposure to contrast media while treatment with simvastatin reversed the effect of contrast media significantly in a dose dependent-manner [24] and agree with Cusumano et al study (2015), who was demonstrated that high dose of atorvastatin has protective effect against inflammation and oxidative stress in ischemia-reperfusion injury of kidney in animal model, also he reported that atorvastatin increase antioxidant enzymes activity of glutathione peroxidase and catalase and inhibit myeloperoxidase [25] and consist with El-Moseelhy et al study (2014), who was reported that treatment with atorvastatin significantly increased GSH and catalase levels, compared with doxorubicin group which cause renal toxicity [26].

In the present study, serum MDA levels significantly raised after exposure to contrast media in CIN group, while treatment with atorvastatin significantly reversed the elevated levels of this marker and reduced it by 33% and 51% in low and high dose respectively, compared with CIN group. These results match with wang et al study (2017), who was reported that serum MDA level markedly increased in contrast induced acute kidney injury group, while treatment with atorvastatin significantly reversed the elevated levels of this marker and reduced it by 33% and 51% in low and high dose respectively, compared with CIN group.

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The main hypothesis of pathogenesis in CIN is local ischemia-reperfusion renal tubules. Interleukin-18 (IL-18) expressed in proximal tubule and activated by caspase-1 during renal ischemia. So, IL-18 can be used for diagnosis of ischemic renal injury in CIN during 24-48 hours [30]. Interleukin-18 is a pro-inflammatory cytokine, its a member of IL-1 cytokine superfamily, describe as important regulator for acquired and innate immune response. It is expressed or induced in autoimmune diseases, sites of acute and chronic inflammation, and infectious diseases [31].

In the current study, serum IL-18 levels were markedly elevated in CIN group compared to control, while its level significantly reduced in low and high dose atorvastatin group by 12% and 26% respectively, compared with CIN group. These results agree with Zhao Kai et al study (2015), who had found that Dongchongxiacao (a herbal Chinese with antioxidant effects) decrease urinary IL-18 levels in CIN by attenuating renal microvessels injury through activating superoxide dismutase[32] and disagree with Jin Ha Park et al study (2016), who had demonstrated that atorvastatin couldn’t lower the incidence of acute kidney injury and serum IL-18 levels in patient undergo valvular heart surgery[33].

Matrix metalloproteinase (MMP) is family of enzymes involved in degradation of the extracellular matrix, it is always present in human tissues in which inflammation occur. The member of MMPs have different roles in normal and pathological inflammatory developments, it act during inflammation to activate cytokines and chemokines and generate of chemokine gradients [34]. WK Han et al (2008) reported that urinary levels of MMP-9 can be used to detect acute renal injury before serum creatinine elevation, he found that urinary MMP-9 concentration in patients with acute kidney injury was significantly higher than patient with chronic renal disease [35]. Also, Van der Zijl et al (2010) demonstrated that levels of MMP-9 in diabetic patients related to the stage of diabetic nephropathy, compared with normal human[36]. In this study, serum MMP-9 levels markedly reduced in low and high doses of atorvastatin by 23.5% and 46% respectively, compared with CIN group. These results were similar to Yao et al results (2010), who reported that simvastatin have potential therapeutic effect in diabetic nephropathy of animal model by down regulating the expression of MMP-9 in renal tissues [37].

The anti-inflammatory properties of atorvastatin depend on the Ras prenylation inhibition. Ras inhibition reduce nuclear factor kappa-B (NFκB) activity involved in various inflammatory pathways and has a central role in atherosclerotic pathology. Atorvastatin reduced the activity of dimer for c-Jun, activator protein 1, and transcriptional factor which have a central role in regulating genes of MMPs, adhesion molecules, endothelial inflammatory response, inducible nitric oxide synthase, chemokines, cytokines, and Fas ligand. Also, atorvastatin induce hypoxia inducible factor 1α (HIF1α), which was decreased by TNF-α in endothelial cells [38].

Conclusion
As conclusion, atorvastatin mechanism for protection against CIN in a dose-dependent manner can mediate by anti-inflammatory and antioxidant effects.

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